

Intrastrand 2'β Hydrogen Abstraction of 5'-Adjacent Deoxyguanosine by Deoxyuridin-5-yl in Z-form DNA

Kiyohiko Kawai and Isao Saito*

Department of Synthetic Chemistry and Biological Chemistry, Faculty of Engineering, Kyoto University, CREST, Kyoto 606-8501, JAPAN

Etsuko Kawashima and Yoshiharu Ishido

Laboratory of Pharmaceutical Chemistry, Tokyo University of Pharmacy and Life Science, Tokyo 192-0392, JAPAN

Hiroshi Sugiyama*

Institute for Medical and Dental Engineering, Tokyo Medical and Dental University, Tokyo 101-0062, JAPAN

Received 24 December 1998; revised 28 January 1999; accepted 29 January 1999

Abstract: In order to identify the site of H abstraction by the deoxyuridin-5-yl in Z-form DNA, we examined the photoreaction of $d(CGCG(2'\beta-D)^{I}UGCG)$ **2-d**/ $d(Cm^{8}GCACm^{8}GCG)$ **3** whose G_{4} residue was deuterated with the diastereoselectivity of 2'R: 2'S=91: 9. Electrospray MS analysis of ribonuclease T1 digested fragments of the $2'\alpha$ -hydroxylation product **4** demonstrated that $2'\alpha$ -hydroxylation results from the abstraction of the $2'\beta$ -H atom by the deoxyuridin-5-yl in Z-form DNA. © 1999 Elsevier Science Ltd. All rights reserved.

DNA local conformations such as A form, Z form, triplex and DNA-RNA hybrid have been suggested to play an important role in biological processes. However, the biological relevance of DNA local conformations is still not well understood due to the lack of a reliable detection method which is applicable to a living cell. Since the DNA local structures are assumed to appear in a very short period of time, utilization of a photoreaction as a DNA conformational probe would provide important structural information on a short-lived DNA local structure. Recently, we found that the 2'-deoxyuridin-5-yl generated in the fixed geometry of a DNA duplex undergoes H abstraction in a highly DNA conformation-dependent manner. For example, the competitive 1'- and 2' α -H abstractions occurred in the B-form DNA, whereas the predominant 1'-H abstraction occurred in a DNA-RNA hybrid. More recently, we found that preferential 2' α -hydroxylation occurred efficiently in Z-form DNA. Inspection of the X-ray structure of the purine-pyrimidine step in Z-DNA indicated that the 2' β H of purine nucleoside is close to the C5 position of the pyrimidine nucleoside, suggesting a selective abstraction of 2' β -H by the deoxyuridin-5-yl (Figure 1). In this study, we examined the photoreaction of d(CGCG(2' β -D)^IUGCG) 2-d/ d(Cm⁸GCA)^{Cm}GCG) 3 in which 2' β -H of the G₄ residue was stereoselectively deuterated, and demonstrated that the 2' α -hydroxylation results from the

abstraction of the 2'β-H by the deoxyuridin-5-yl.

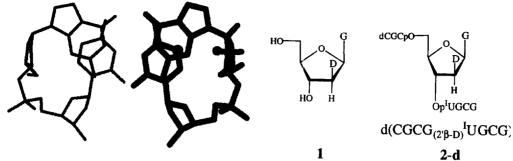


Figure 1. Purine-pyrimidine step of Z-DNA. $C2^{\circ}\beta$ hydrogen and C5 carbon are represented by balls. The complementary strand is shown as a thin line.

The stereoselectively deuterated nucleoside, $(2'S)-\{2'-2H\}-2'-\text{deoxyguanosine}$ 1 with the diastereoselectivity of 2'R: 2'S=91: 9, was synthesized according to the previous method⁶ and incorporated into the oligonucleotide via the standard phosphoramidite method. The octanucleotide $d(Cm^8GCACm^8GCG)$ 3 as a complement for $d(CGCG^IUGCG)$ 2 and $d(CGCG(2'\beta-D)^IUGCG)$ 2-d was prepared as previously reported.^{4,7} The structure of the octamers was confirmed by enzymatic digestion and by electrospray MS (ESMS). The 5-iodouracil-containing Z-form duplex 2-d/3 was irradiated at 302 nm with a transilluminator. The $2'\alpha$ -hydroxylated product 4 produced was isolated and treated with ribonuclease T1. As previously reported,⁴ the $2'\alpha$ -hydroxylated product 4 was quantitatively hydrolyzed to d(CGC)rG>(>: cyclic phosphate) 5 and d(UGCG) 6 (Scheme 1).

Scheme 1

In order to identify the position of deuterium (D) after the $2'\alpha$ -hydroxylation reaction, the hydrolyzed fragments 5 and 6 were subjected to ESMS. Figure 2 shows the ESMS of 5 and 6 obtained from the photoreaction of 2-d/3 and the unlabeled octamers (control), indicating that $2'\beta$ -D is specifically abstracted and incorporated into 6. These results clearly indicate that $2'\beta$ -H of G_4 was abstracted by the 5'-adjacent deoxyuridin-5-yl radical as an initiation step for the formation of the $2'\alpha$ -hydroxylated product 4 (Scheme 2). The intramolecular kinetic isotope effects (KIE) $[(k_H/k_D)=(\%4/\%4-d)]$ obtained for the $2'\alpha$ -hydroxylation by photoirradiation of the IU-containing Z-form oligonucleotide was found to be 1.2 ± 0.1 . The magnitude of KIE obtained in the present study was in the range of 1.0 to 5.8 which were previously reported for 1', 4',

and 5' H abstraction by several antitumor antibiotics. However, the KIE value of 1.2±0.1 is substantially lower than the value 7.2 which was observed for the 2'α-H abstraction in B-form DNA.^{2c}

Scheme 2

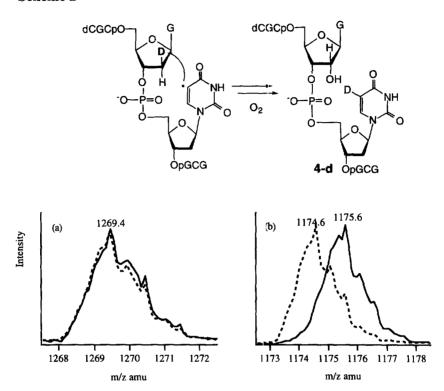


Figure 2. ESMS of (a) d(CGC)rG> 5 and (b) d(UGCG) 6. MS of the products obtained from the deuterated and unlabeled octamers are indicated by the solid and dashed lines, respectively.

We previously developed a simple method to evaluate the conformational energy needed to achieve the transition state for H abstraction in DNA.³ The calculation provided a qualitative explanation for the competitive 1'- and 2' α -H abstractions and the selective 1'-H abstraction by the deoxyuridin-5-yl in DNA duplex and DNA-RNA hybrid, respectively. Based on this method, the transition states for the 1'-, 2' α - and 2' β -H abstractions in Z-form DNA were calculated and are summarized in Table 1. These results suggest that the AMBER energy of the 2' β H-abstraction in Z-form DNA is significantly low compared to the other transition states. The smaller KIE observed in Z-DNA would be a consequence of the lower energy for 2' β -H abstraction in Z-form DNA. In summary, using stereoselectively deuterated octanucleotide, we demonstrated that the C2' α -hydroxylation was a consequence of the reaction of O₂ with the C2' radical which was resulted from 2' β -H abstraction by the deoxyuridin-5-yl formed in Z-form DNA.

Table 1. Minimized AMBER Total Energies (kcal/mol) of Octamer Containing Transition States for Deoxyribose Hydrogen Abstraction by Deoxyuridin-5-yl

Transition state .				
Conformation	C1'	C2'α	C2'β	Unmodified
Z-form	-91.1 (14.6) ^b	-90.7 (15.0) ^b	-95.2 (10.5) ^b	-105.7

^{*}Starting structures were generated from standard Z DNA in a builder module of Insight II (MSI, San Diego, CA). The models were energy minimized by conjugate gradient (0.001 kcal/molÅ). To prevent unusual distortion of the helix, the terminal residues were tethered to the initial structures (100 kcal/molÅ). Total energies of the molecules were evaluated using AMBER force field and distance-dependent dielectric of 4r. bThe value in parentheses is the difference in energy from the corresponding unmodified octamers d(CGCGUGCG)/d(CGCACGCG).

References and Notes

- (a) Cozzarelli, N. R.; Wang, J. C. DNA Topology and Its Biological Effects 1990, Cold Spring Harbor Laboratory Press: New York. (b) Sinden, R. R. DNA Structure and Function 1994, Academic Press: New York. (c) Travers, A. A. Annu. Rev. Biochem. 1989, 58, 427.
- (a) Sugiyama, H.; Tsutsumi, Y.; Saito, I. J. Am. Chem. Soc. 1990, 112, 6720.
 (b) Sugiyama, H.; Tsutsumi, Y.; Fujimoto, K.; Saito, I. J. Am. Chem. Soc. 1993, 115, 4443.
 (c) Sugiyama, H.; Fujimoto, K.; Saito, I.; Kawashima, E.; Sekine, T.; Ishido, Y. Tetrahedron Lett. 1996, 37, 1805.
- 3. Sugiyama, H.; Fujimoto, K.; Saito, I. Tetrahedron Lett. 1997, 38, 8057.
- 4. Kawai, K.; Saito, I.; Sugiyama, H. J. Am. Chem. Soc. in press.
- (a) Wang, A. H.-J.; Quigley, G. J.; Kolpak, F. J.; Crawford, J. L.; van Boom, J. H.; van der Marel, G. A.; Rich, A. Nature 1979, 282, 680-686.
 (b) Rich, A.; Nordheim, A.; Wang, A. H.-J. Ann. Rev. Biochem. 1984, 53, 791. (c) Rich, A. Ann. N. Y. Acad. Sci. 1994, 726, 1.
- Kawashima, E.; Aoyama, Y.; Sekine, T.; Miyahara, M.; Radwan, M. F.; Nakamura, E.; Kainosho, M.;
 Kyogoku, Y.; Ishido, Y. J. Org. Chem. 1995, 60, 6980-6986. (b) Kawashima, E.; Sekine, T.;
 Ishido, Y. Nucleoside and Nucleotides 1995, 14, 333.
- 7. Sugiyama, H.; Kawai, K.; Matsunaga, A.; Fujimoto, K.; Saito, I.; Robinson, H.; Wang, A. H. -J. *Nucleic Acids Res.* **1996**, *24*, 1272-1278.
- (a) Kozarich, J. W.; Worth, L.; Frank, B. L.; Christner, D. F.; Vanderwall, D. E.; Stubbe, J. Science 1989, 245, 1396. (b) Kappen, L. S.; Goldberg, I. H.; Wu, S. H.; Stubbe, J.; Worth, L.; Kozarich, J. W. J. Am. Chem. Soc, 1990, 112, 2797. (c) Frank, B. L.; Worth, L.; Christner, D. F.; Kozarich, J. W.; Stubbe, J.; Kappen, L.; Goldberg, I. H. J. Am. Chem. Soc, 1991, 113, 2271. (d) Kappen, L.; Goldberg, I. H.; Frank, B. L.; Worth, L.; Christner, D. F.; Kozarich, J. W.; Stubbe, J. Biochemistry 1991, 30, 2034. (e) Hangeland, J. J.; De Voss, J. J.; Heath, J. A.; Townsend, C. A.; Ding, W.; Ashcroft, J. S.; Ellestad, G. A. J. Am. Chem. Soc, 1992, 114, 9200. (f) Absalon, M. J.; Wu, W.; Kozarich, J. W.; Stubbe, J. Biochemistry, 1995, 34, 2076. (g) Greenberg, M. M.; Barvian, M. R.; Cook, G. P.; Goodman, B. K.; Matray, T. J.; Tronche, C.; Venkatesan, H. J. Am. Chem. Soc, 1997, 119, 1828.